

Applied Biostatistics

<https://moodle.epfl.ch/course/view.php?id=15590>

- Research process
- Basic experimental design ideas
- Analysis of variance
- ANOVA modeling with R

Research process

- Scientific *question* of interest
- Decide *what data* to collect (and how)
- Collection and *analysis* of data
- Conclusions, generalizations : *inference* on the population
- *Communication* and dissemination of results

Generic question : Does a 'treatment' have an 'effect' ?

Examples :

- Does smoking cause cancer, heart disease, etc ?
- Does eating oat bran lower cholesterol ?
- Does echinacea prevent illness ?
- Does exercise slow the aging process ?

Approach the question :

- One simple method for resolving this type of question is to *compare two groups* of study subjects :
 - *Control group* : gives a base level for comparison
 - *Treatment group* : group receiving the 'treatment'

Types of studies

- A basic means to address this type of question involves *comparing two groups* of study subjects :
 - *Control group* : provides a baseline for comparison
 - *Treatment group* : group receiving the 'treatment'
- *Experimental study* : subjects assigned to groups by the investigator
 - *randomization* : protects against bias in assignment to groups
 - *'blind', 'double-blind'* : protects against bias in outcome assessment/measurement
 - *placebo* : artificial/fake treatment
- *Observational study* : subjects 'assign' themselves to groups
 - *confounder* : associated with both group membership/risk factor *and* with the outcome of interest

A few comments

- With a well-planned and well executed controlled experiment, it is possible to infer *causality*
- This is *not possible* with observational studies due to the presence of confounders
- With confounding, it is not possible to tell whether the observed difference between groups is due to the *treatment* or to the *confounding factor*
- Not always possible to carry out an experiment, for *practical* and *ethical* reasons

Example : Hibernation

- General question : *How do changes in an animal's environment induce hibernation ?*
- What changes should be studied ??
 - temperature
 - photoperiod (daylight duration)
- What measures to take ?
 - nerve enzymatic activity (Na⁺K⁺ATP-ase)
- What animal to study ?
 - golden hamster, 2 organs

Specific question

- General question : *How do changes in an animal's environment induce hibernation ?*
- *Specific question* : What is the effect of changing daylight duration on the enzyme concentration of the sodium pump in two golden hamster organs ?

Sources of variability

- Variability due to the conditions of interest (wanted)
 - Duration (long or short)
 - Organ (heart or brain)
- Variability of the response (NOT wanted) : measurement error
 - Preparation of the enzyme suspension
 - Instrument calibration/standardization
- Variability in experimental units (NOT wanted)
 - biological differences between hamsters
 - environmental differences

Types of variability

- Systematic, expected (wanted)
- Random variation (can manage this)
- Systematic, unexpected (NOT wanted)
 - biased results
 - e.g., what time the measurements are made

Questions for the hibernation study

- *Long or short* : Is there an effect of daylight duration on enzyme concentration ?
- *Heart vs. Brain* : Are the concentrations different in the 2 organs ?
- *Interaction* : Is the *difference* in enzyme concentration (long/short) *different* for heart and brain ?
- *Hamsters* : Variability between hamsters ?
- *Measurement error* : What is the error due to the measurement process for enzyme concentration ?

Experimental design – why do we care?

- Poor design *costs* :
 - time, money, ethical considerations
- To ensure *relevant data* are collected, and can be analyzed to test the scientific hypothesis/ question of interest
 - Decide *in advance* how data will be analyzed
 - 'Designing the experiment' = 'Planning the analysis'
- **The design is about the biology**

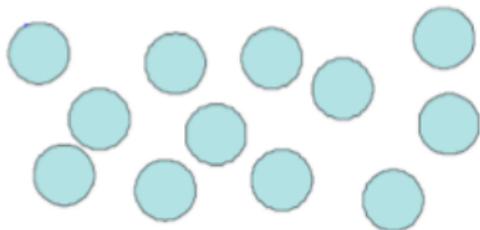
Common experimental designs

- *Completely randomized design* (CRD)
 - compares 2 (or more) *levels* of a single *factor*
 - analysis : 1-way anova (below)
- *Randomized Block Design* (RBD)
 - compares 2 (or more) *levels* of a single *factor*
 - observations in *blocks*
 - analysis : similar to unreplicated 2-way anova
- *(Full) Factorial design*
 - levels from multiple factors varied and studied *simultaneously*
 - can detect *interaction* between factors
 - analysis : 2-way (or multi-way) anova

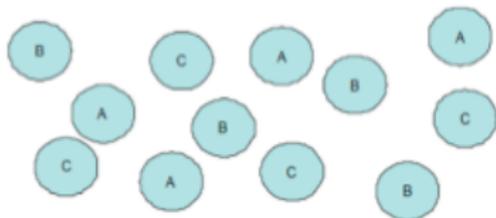
Completely randomized experiment

- Study subjects (experimental units) *homogeneous*
- *Randomized* to treatments (factor levels)

experimental units



randomized to treatments

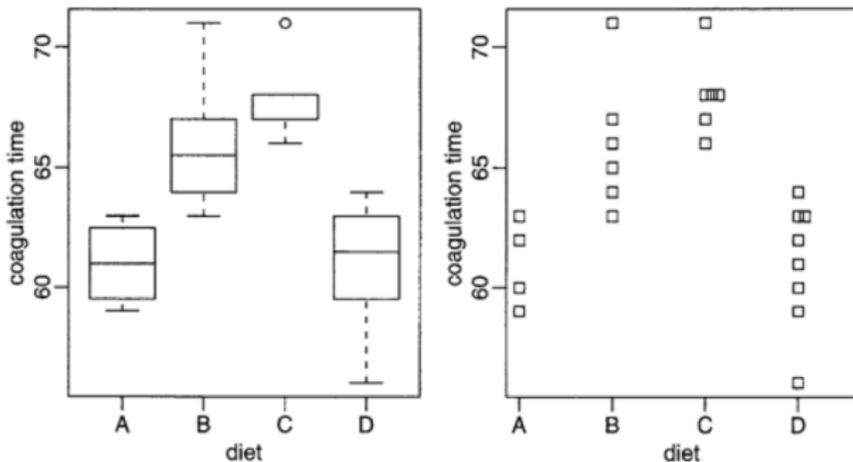


Data example : Blood coagulation time

- 24 animals
- Randomly assigned
- to 4 different diets
- Measured blood coagulation times from samples taken in a random order
- *As always*, the first step of analysis is **EXPLORATORY**

Compare distributions with boxplots

- We hope we *don't* see :
 - *outliers* – points outside the whiskers
 - *skewness* – asymmetrical boxes
 - *unequal variance* – clearly unequal box sizes
- **BUT** : don't over-interpret boxplots based on small n



Trees

- A study is conducted to investigate the growth of a certain type of tree at an elevation of 675 meters
- The variable of interest is the core measurement (in cm) for a 10 year period
- The theory is that the mean should be at least 1.75
- In a random sample of 10 measurements, the mean was 2 with an SD of 0.5

Hypothesis test for this setup

- Identify the population parameter being tested
Here, the parameter being tested is the population mean core measurement μ
- Formulate the NULL and ALT hypotheses
 $H: \mu = 1.75$ (or $\mu \leq 1.75$)
 $A: \mu > 1.75$
- Compute the Test Statistic (TS)
 $t = (2 - 1.75) / (.5 / \sqrt{10}) = 1.58$
- Compute the p -value
Here, $p = P(t_9 > 1.58) = 0.07$
- **Decision Rule** : REJECT H if the p -value $\leq \alpha$
If we use $\alpha = 0.05$, the decision here will be DO NOT REJECT H (but just barely!)

More trees

- Now say we are interested in whether the mean core measurement is the same in trees at 675 meters and trees at 825 meters
- Assume that we have a random sample also of size 10 of trees at 825 meters, with a mean core measurement of 2.65 cm and SD 1.15 cm
- How might we test the null that the means are the same, against the alternative that they are different?

Test for comparing two (independent) means : equal variances

- We want to compare the means of two sets of measures :
 - Group 1 (p. ex. 'control') : x_1, \dots, x_n
 - Group 2 (p. ex. 'treatment') : y_1, \dots, y_m

- We can *model* these data as :

$$x_i = \mu + \epsilon_i; i = 1, \dots, n;$$

$$y_j = \mu + \Delta + \tau_j; j = 1, \dots, m,$$

where Δ signifies the effect of the treatment (compared to the 'control' group)

- $H: \Delta = 0$ vs. $A: \Delta \neq 0$ or $A: \Delta > 0$ or $A: \Delta < 0$

Equal variances, cont.

$$\blacksquare T = \text{obs. diff.} / \text{ES}(\text{obs. diff.}) = \frac{\Delta}{\sqrt{\text{Var}(\hat{\Delta})}};$$

$$\hat{\Delta} = \bar{y} - \bar{x}; \text{Var}(\hat{\Delta}) = \frac{\sigma^2}{n} + \frac{\sigma^2}{m} = \frac{n+m}{nm} \sigma^2$$

- We assume that :

- the variances of the 2 samples are *equal* :

$$\text{Var}(\epsilon) = \text{Var}(\tau)$$

- the observations are *independent*

- *the 2 samples are independent*

- We can estimate the variances *separately* :

$$s_x^2 = ((x_1 - \bar{x})^2 + \dots + (x_n - \bar{x})^2) / (n - 1)$$

$$s_y^2 = ((y_1 - \bar{y})^2 + \dots + (y_m - \bar{y})^2) / (m - 1)$$

- When the variances are *equal*, we can combine the two estimators : $s_p^2 = ((n - 1)s_x^2 + (m - 1)s_y^2) / (n + m - 2)$

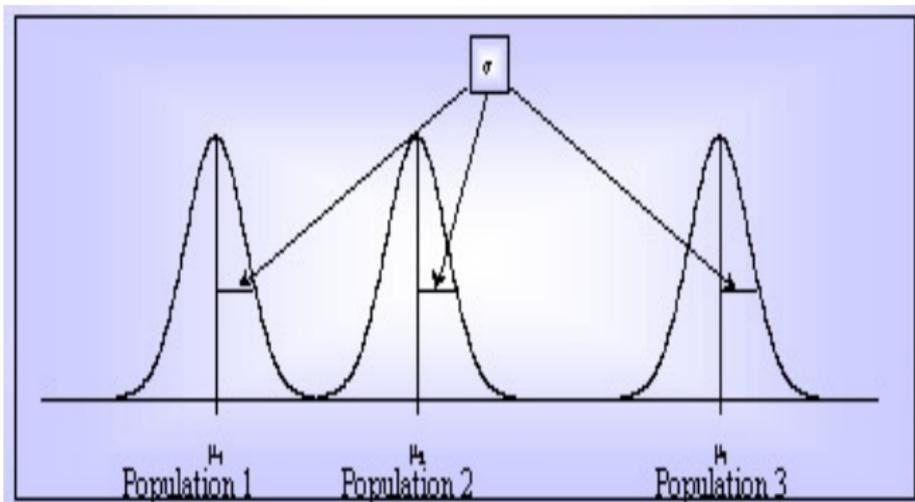
$$\Rightarrow t_{obs} = \frac{\bar{y} - \bar{x}}{\sqrt{s_p^2(n + m) / (nm)}} \sim t_{n+m-2} \text{ under } H$$

Trees one more time!

- You guessed it! Now say we are also interested in trees at 975 meters as well
- Want to make a *three-way* comparison
- Have a random sample (size 10 again) and find the mean is 2.5 and the SD is 1
- How might we test the null that all three means are the same, against the alternative that at least one is different?

ANOVA

- Abbreviation for *AN*alysis *Of* *VA*riance (analyse de variance)
- But it's a test for a difference in *means*
- The idea :



Principle

- The variation (total sum of squared deviations) consists of *2 components*
 - *individual* fluctuations : variability *intra-group* (error)
 - *between group* fluctuations : variability *inter-group* (treatment)
- Variability inter-group > Variability intra-group
⇒ (at least) 2 means are (significantly) different
- General principle :
 - *Decompose* the total sum of squared deviations into its 2 (orthogonal) parts
 - *Test* if the MS_{inter} (MSB) is (significantly) bigger than the MS_{intra} (MSW, or MSE)

Hypothesis tests

- Notation :
 - k groups
 - n_i individuals in group i
 - observations x_{ij} (observation j from group i)
- $H: \mu_1 = \mu_2 = \dots = \mu_k$
 $A: \exists \mu_i \neq \mu_j$ (at least 1 mean is different from the others)
- ANOVA is a rather *robust* test (resultats not too influenced by small deviations from the assumptions)

Pairs of tests : why not ?

Why not start off by carrying out tests (z or t) for each pair of samples ?

- For m comparisons (independent), the probability of rejecting at least one H can be expressed as $\alpha_m = 1 - (1 - \alpha)^m$; now for $\alpha = 0.05$:
- 3 tests \implies Type I error = 0.14
- 5 tests \implies Type I error = 0.23
- 10 tests \implies Type I error = 0.4
- 21 tests \implies Type I error = 0.66

\implies Type I error no longer controlled at level $\alpha = 0.05$
(anti-conservative/liberal test)

The models

- $\epsilon_{ij} \sim \text{iid } N(0, \sigma^2)$
- Under H , the model is :

$$x_{ij} = \mu + \epsilon_{ij}$$

- Under A , the model is :

$$x_{ij} = \mu + \alpha_i + \epsilon_{ij},$$

where α_i is *the effect of modality/level i* of facteur A on the variable X

- For each model, we can derive an estimator for the residual variance

Sum of squares

- Goal : test difference between means of two (or more) groups
 - Between SS measures the difference
- The difference must be measured relative to the variance within the groups
 - Within SS
- *F-test* : considers the ratio of B/W
- The larger F is, the more significant the difference

The ANOVA procedure

- Subdivide observed total sum of squares into several components
- Pick appropriate significance point for a chosen Type I error from an F table
- Compare the observed components to test the NULL hypothesis

Parameter estimation

- Under $H : x_{ij} = \mu + \epsilon_{ij} :$

$$\hat{\mu} = \bar{x} = \frac{1}{n} \sum_{i=1}^k, \sum_{j=1}^{n_i} x_{ij}, \quad n = \sum_{i=1}^k n_i$$

- Under $A : x_{ij} = \mu + \alpha_i + \epsilon_{ij} :$

$$\hat{\mu} + \hat{\alpha}_i = \bar{x}_i = \frac{1}{n_i} \sum_{j=1}^{n_i} x_{ij}, \quad n = \sum_{i=1}^k n_i,$$

which gives us $\hat{\alpha}_i = \bar{x}_i - \bar{x}$

$$\hat{\epsilon}_{ij} = x_{ij} - \hat{x}_{ij} = x_{ij} - \hat{\mu} - \hat{\alpha}_i = x_{ij} - \bar{x} - (\bar{x}_i - \bar{x}) = x_{ij} - \bar{x}_i$$

Decomposition of the total variation

- The model under A : $x_{ij} = \mu + \alpha_i + \epsilon_{ij}$
- with estimators : $x_{ij} = \bar{x} + (\bar{x}_i - \bar{x}) + (x_{ij} - \bar{x}_i)$
- $\implies (x_{ij} - \bar{x}) = (\bar{x}_i - \bar{x}) + (x_{ij} - \bar{x}_i)$
- with sum of squares :
- $(x_{ij} - \bar{x})^2 = (\bar{x}_i - \bar{x})^2 + (x_{ij} - \bar{x}_i)^2 + 2(\bar{x}_i - \bar{x})(x_{ij} - \bar{x}_i)$
- and sums for individuals (j) :

$$\sum_{j=1}^{n_i} (x_{ij} - \bar{x})^2 =$$

$$n_i (\bar{x}_i - \bar{x})^2 + \sum_{j=1}^{n_i} (x_{ij} - \bar{x}_i)^2 + 2(\bar{x}_i - \bar{x}) \sum_{j=1}^{n_i} (x_{ij} - \bar{x}_i)$$

Decomposition, cont.

- Thus, $2(\bar{x}_i\bar{x}) \sum_{j=1}^{n_i} (x_{ij} - \bar{x}_i) = 0$, since $\sum_{j=1}^{n_i} (x_{ij} - \bar{x}_i) = 0$ ($E[\epsilon_{ij}] = 0$)
- Therefore,

$$\sum_{j=1}^{n_i} (x_{ij} - \bar{x})^2 = n_i (\bar{x}_i - \bar{x})^2 + \sum_{j=1}^{n_i} (x_{ij} - \bar{x}_i)^2$$

- with the sums for the factor levels :

$$\sum_{i=1}^k \sum_{j=1}^{n_i} (x_{ij} - \bar{x})^2 + \sum_{i=1}^k n_i (\bar{x}_i - \bar{x})^2 + \sum_{i=1}^k \sum_{j=1}^{n_i} (x_{ij} - \bar{x}_i)^2$$

$$\implies SS_{total} = SS_{groups} + SS_{error}$$

Test principle

- 1-factor analysis of variance tests the effect of one *factor* A having k modalities on the means of a *quantitative variable* X
- The tested hypotheses are :
 $H : \mu_1 = \mu_2 = \dots = \mu_k = \mu$ vs. $A : \exists \mu_i \neq \mu_j$
- Test if the ratio of 2 variance estimators is close to 1
- The variance estimators associated are :
 - *Total variance* : $SS_{total}/(n-1)$
 - *Variance due to factor A* (MS_{trts}) : $SS_{trts}/(k-1)$
 \implies estimator of σ^2 if H is true
 - *Residual variance* (MS_{error}) : $SS_{error}/(n-k)$
 \implies estimator of σ^2 whichever model

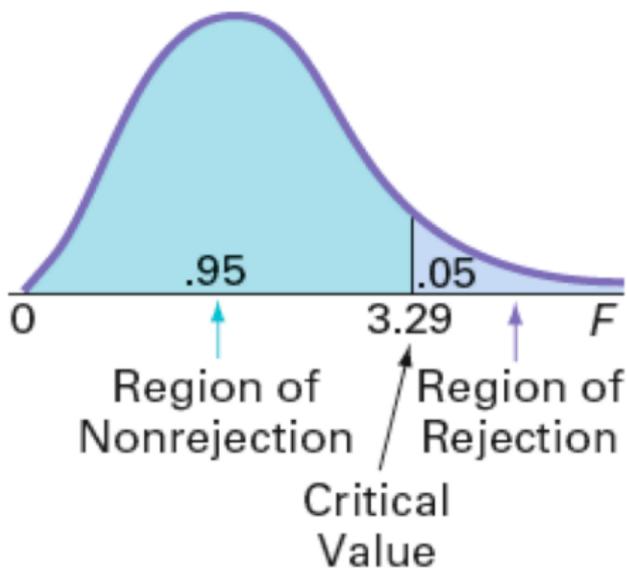
Test statistic

- Under H , $SS_{trts}/(k-1)$ and $SS_{error}/(n-k)$
 \Rightarrow *estimators of the same parameter σ^2*
- Thus (under H), the ratio $\frac{SS_{trts}/(k-1)}{SS_{error}/(n-k)} \approx 1$
- Under A , at least 1 $\alpha_i \neq 0$ and $SS_{error}/(n-k)$ is a unique estimator of σ^2 ; $SS_{trts}/(k-1) \gg SS_{error}/(n-k)$
- Thus (under A), the ratio $\frac{SS_{trts}/(k-1)}{SS_{error}/(n-k)}$ *much larger than 1*
- \Rightarrow F -Test *unilateral* in every case
- $F_{obs} = \frac{SS_{trts}/(k-1)}{SS_{error}/(n-k)} = MS_{trts}/MS_{error}$
- Test statistic is distributed according to a Fisher F distribution, with $k-1$ (num) and $n-k$ (denom) degrees of liberty (df)

ANOVA table

ANOVA table

source	df	SS	MS (=SS/df)	F	p-value
treatments	$k - 1$	SS_{trts}	$SS_{trts}/(k - 1)$	MS_{trts}/MS_{error}	$P(F_{obs} > F_{k-1, n-k})$
error	$n - k$	SS_{error}	$SS_{error}/(n - k) (= \hat{\sigma}^2)$		
total (corr.)	$n - 1$	SS_{total}			



What does it mean when we reject H ?

- The null hypothesis H is a joint (global) one : that *all* the population means are equal
- When we reject the null hypothesis, that does not mean that all the means are different !!
- It means that *at least one* is different
- To know which is different, we can carry out '*post hoc*'/*a posteriori* tests (pairs of tests, for example – below)

Model formulas in R

- A simple *model formula* in R looks something like :
`yvar ~ xvar1 + xvar2 + xvar3`
- We could write this model (algebraically) as

$$y = \beta_0 + \beta_1 x_1 + \beta_2 x_2 + \beta_3 x_3 + \epsilon$$

- By default, an intercept is included in the model – you *don't* have to include a term in the model formula
- If you want to leave the intercept out :
`yvar ~ -1 + xvar1 + xvar2 + xvar3`

More on model formulas

- We can also include interaction terms in a model formula :
 $yvar \sim xvar1 + xvar2 + xvar3$
- Examples :
 - $yvar \sim xvar1 + xvar2 + xvar3 + xvar1 : xvar2$
 - $yvar \sim (xvar1 + xvar2 + xvar3)^2$
 - $yvar \sim (xvar1 * xvar2 * xvar3)$

More on model formulas

- The generic form is **response** ~ **predictors**
- The predictors can be **numeric** or **factor**
- Other symbols to create formulas with *combinations of variables* (e.g. *interactions*)
 - **+** to *add* more variables
 - **-** to *leave out* variables
 - **:** to introduce *interactions* between two (or more) terms
 - ***** to include *both the interactions and all lower order terms* (**a*b** is the same as **a+b+a:b**)
 - **^ n** *adds all terms* including interactions up to order *n*
 - **I()** treats what's inside () as a *mathematical expression*

Tables of group means for chicks data

		Groundnut	Soybean	Mean
Level of protein	0	6876	7452	7064
	1	6893	6961	7927
	2	6719	6624	6671
Mean		6763	7012	6887

		G-nut	Soy	Level of protein			Mean
				0	1	2	
Level of fish	0	6537	6752	6750	6595	6588	6644
	1	6989	7273	7379	7259	6755	7131
Mean		6763	7012	7064	6927	6671	6887

Interpreting R output

```
> chicks.aov <- aov(Weight ~ House + Protein*LP*LS)
> summary(chicks.aov)
```

	Df	Sum Sq	Mean Sq	F value	Pr(>F)	
House	1	708297	708297	15.8153	0.0021705	**
Protein	1	373751	373751	8.3454	0.0147366	*
LP	2	636283	318141	7.1037	0.0104535	*
LS	1	1421553	1421553	31.7414	0.0001524	***
Protein:LP	2	858158	429079	9.5808	0.0038964	**
Protein:LS	1	7176	7176	0.1602	0.6966078	
LP:LS	2	308888	154444	3.4485	0.0687641	.
Protein:LP:LS	2	50128	25064	0.5596	0.5868633	
Residuals	11	492640	44785			

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

R output for the coagulation example

```
> g <- lm(coag ~ diet, data=coagulation)
> summary(g)
```

Call:

```
lm(formula = coag ~ diet, data = coagulation)
```

Residuals:

Min	1Q	Median	3Q	Max
-5.00	-1.25	0.00	1.25	5.00

Coefficients:

	Estimate	Std. Error	t value	Pr(> t)	
(Intercept)	6.100e+01	1.183e+00	51.554	< 2e-16	***
dietB	5.000e+00	1.528e+00	3.273	0.003803	**
dietC	7.000e+00	1.528e+00	4.583	0.000181	***
dietD	2.991e-15	1.449e+00	0.000	1.000000	

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Residual standard error: 2.366 on 20 degrees of freedom

Multiple R-squared: 0.6706, Adjusted R-squared: 0.6212

F-statistic: 13.57 on 3 and 20 DF, p-value: 4.658e-05

ASSUMPTIONS

- *Independence* : The k groups (samples) are independent, as well as the individuals within groups; the ensemble of the n individuals are placed *at random* (*randomization*) between the k modalities for the controlled factor A , with n_i individuals receiving treatment i .
- *Homoscedasticity* : The k populations have the same variance; the factor A acts only on the *mean* of the variable X and does not change its variance
- *Normality* : The variable studied follows a Normal distribution in the k populations compared (or the CLT applied to the means if the n_i are 'sufficiently large')

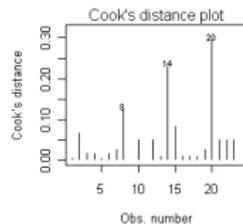
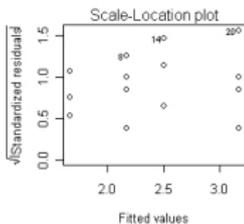
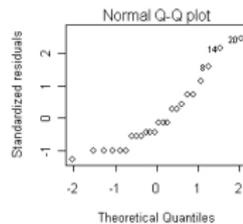
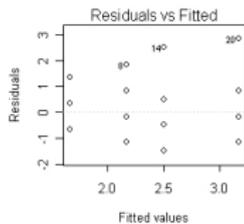
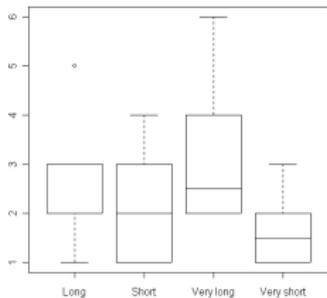
Model assessment : Normality

- Boxplots of observations (or residuals) should be rather symmetric
- A graph of the sample mean vs. variaces should not display any pattern
- QQ-plot (normal) plot of the observations (or residuals) should form a straight line
- Check whether there are any unusual or influential values

Model evaluation : Homogeneity of variance

- Boxplots of the observations should show similar variability
- Variability of the residuals should be similar in the graph of residuals versus fitted values
- It is also possible to carry out formal hypothesis tests (e.g. Bartlett, Levene), but these are not useful for diagnosing problems

Some diagnostic plots



Evaluation of the model : Independence

- Graphics : residuals vs. group mean, might indicated autocorrelation for example
- Normally, treat the question of independence during the conception of the experiment, for example using randomization or perhaps other methods

ANOVA : after the test

- Once all the conditions for an ANOVA have been verified and the analysis carried out, two conclusions are possible :
 - we reject H
 - we do not have enough evidence to reject H
- If H is not rejected, we conclude that there are not significant differences between group means
- If we DO reject H , typically we are interested in *identifying the modalities/factor levels* that are responsible for the significant result

Multiple comparisons

- Comparing means of pairs of treatments
- Carried out *after* a significant ANOVA
- Types of comparisons
 - planned (*a priori*) : independent of the ANOVA results ; the theory predicts which treatments should be different
 - unplanned (*a posteriori*) : the comparisons are decided *based on the ANOVA results*
- $H : \mu_i = \mu_j$ vs. $A : \mu_i \neq \mu_j$
- Test statistic

$$t = \frac{\bar{y}_i - \bar{y}_j}{\sqrt{\hat{\sigma}^2 (1/n_i + 1/n_j)}}$$

- $(\hat{\sigma}^2 = MS_{error}) ; df = df_{error}$

Bonferroni method – global control

- To maintain the global level α_e at level α , we must *adjust α for each comparison by the total number of comparisons*
- In this way, α_e is *independent* of the number of comparisons
- Simplest method : method of Bonferroni

$$\alpha' = \alpha/k,$$

where k = number of comparisons (tests)

- $p_{adj} = \min(kp, 1)$
- Bonferroni's method assures that the global level is *no larger than the desired level*
- (That property makes this method *conservative*, and thus less powerful than other methods, but it is applicable for any situation)

Multiple comparisons : Tukey Honest Significant Differences

- Interested in simultaneous confidence intervals or tests for *differences in the mean outcome X* for *pairs of levels* of a factor
- To test all pairwise comparisons among means using the *Tukey HSD*, calculate HSD for each pair of means :

$$q_s = \frac{M_i - M_j}{\sqrt{MSW/n_{group}}},$$

where M_k is the mean of group k , $M_i > M_j$

- For hypothesis testing, compare q_s to a q value from the *studentized range distribution* (difference between largest and smallest sample means divided by pooled sample SD $\sqrt{2/n}$)
- Reject the null at level α if $q_s > q_\alpha$
- CI : $(\bar{y}_i - \bar{y}_j) \pm \frac{q_{\alpha;k;N-k}}{\sqrt{2}} \hat{\sigma}_e \sqrt{2/n}$; $i, j = 1, \dots, k, i \neq j$
- k = number of populations; N = total sample size

(Complete) Randomized block design

- Assume that the hamsters have come from 4 *different litters*, 2 hamsters per litter
- We expect that hamsters born in the same litter are *more similar* to each other than hamsters from a different litter
- For each pair of hamsters *randomly assign* short or long to one member of each pair
- Example (toss a fair coin, for example) :
S, L // L, S // S, L // S, L
- Analysis : 2-way analysis of variance

Replication, Randomization, Blocking

- These are the 'big three' of experimental design
- **Replication** – to reduce *random variation* of the test statistic ; increases generalizability
- **Randomisation** – to reduce/remove *bias*
- **Blocking** – to reduce *unwanted variation*
- Idea here is that units within a block are similar to each other, but different *between* blocks
- 'Block what you can, randomize what you cannot'

Factorial crossing

- Compare 2 (or more) sets of conditions in the *same experiment*
- Designs with factorial treatment structure allow you to measure *interaction* between two (or more) sets of conditions that influence the response
- Factorial designs may be either *observational* or *experimental*

Interaction

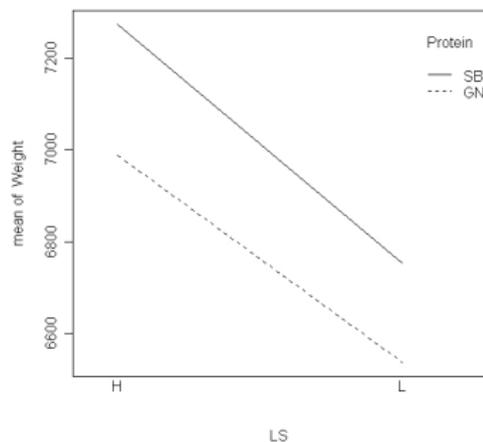
- Interaction is very common (and very important) in science
- Interaction is a *difference of differences*
- Interaction is present if the effect of one factor *is different* for different levels of the other factor
- *Main effects can be difficult to interpret in the presence of interaction*, because the effect of one factor *depends* on the level of the other factor

Factorial experimental design and interaction

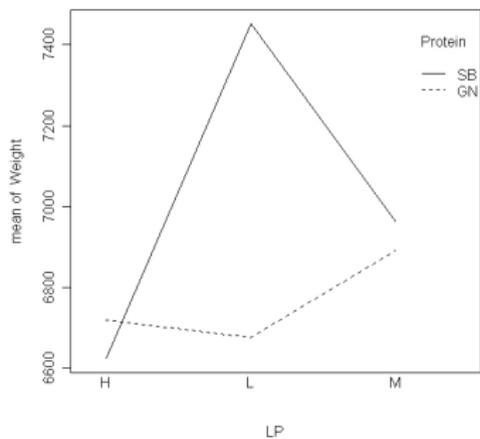
- Example : hibernation study
 - General question : *How do changes in an animal's environment induce hibernation ?*
 - *Specific question* : What is the effect of changing daylight duration on the enzyme concentration of the sodium pump in two golden hamster organs ?
- Compare two (or more) sets of conditions in the *same experiment* : long/ short AND heart/brain
- In this example, there are 4 combinations of conditions :
 - Long/Heart, Long/Brain, Short/Heart, Short/Brain
- Interaction = 'difference of differences'
- There is an *interaction* when the effect of the association of combined treatments is not the sum of treatment effects
- In the case of interaction, the effect of a treatment *varies according to whether it is associated with the other treatment*
- The interpretation of individual effects is more difficult when interaction is present

Interaction plot

pas d'interaction



interaction



Advantages of factorial experiments

- *More efficient* (powerful) than a series of experiments studying one factor at a time
- Permits estimation of *interaction* between sets of conditions that may affect the response
- All data are used for effect estimation

2-way ANOVA

- Simultaneous study of a factor A with I levels and a factor B with J levels
- For each pair of levels (A, B) :
 - we have a sample
 - all samples are of the *same size* n (balanced design)
- Suppositions :
 - the populations studies are Normally distributed
 - the population variances are all equal (homoscedasticity)
 - the samples are taken randomly and independently in the populations

Complete model

- The *complete model* : with interaction
- $y_{ijk} = \mu + \alpha_i + \beta_j + \gamma_{ij} + \epsilon_{ijk}$
- $E[\epsilon_{ijk}] = 0$, $Var(\epsilon_{ijk}) = \sigma^2$, $Cov(\epsilon_{ijk}, \epsilon_{i'j'k'}) = 0$ si $(ijk) \neq (i'j'k')$

ANOVA table

source	df	SS	MS	F
A	$I - 1$	$nJ \sum_{i=1}^I (\bar{y}_{i..} - \bar{y}_{...})^2$	SS_A / df_A	MS_A / MS_{err}
B	$J - 1$	$nI \sum_{j=1}^J (\bar{y}_{.j.} - \bar{y}_{...})^2$	SS_B / df_B	MS_B / MS_{err}
AB	$(I - 1)(J - 1)$	$n \sum_{j=1}^J \sum_{i=1}^I (y_{ij.} - \bar{y}_{i..} - \bar{y}_{.j.} + \bar{y}_{...})^2$	SS_{AB} / df_{AB}	MS_{AB} / MS_{err}
error	$IJ(n - 1)$	$\sum_{k=1}^n \sum_{j=1}^J \sum_{i=1}^I (y_{ijk} - \bar{y}_{ij.})^2$	SS_{err} / df_{err}	
total (corr.)	$nIJ - 1$	$\sum_{k=1}^n \sum_{j=1}^J \sum_{i=1}^I (y_{ijk} - \bar{y}_{...})^2$		

Hypothesis tests

- Test for interaction

$$H: \gamma_{ij} = 0, i = 1, \dots, I, j = 1, \dots, J$$

- Test statistic :

$$F_{AB} = MS_{AB}/MS_{error} \sim F_{(I-1)(J-1), IJ(n-1)} \text{ under } H$$

- Test for effect of factor A

$$H: \alpha_i = 0, i = 1, \dots, I$$

- Test statistic :

$$F_A = MS_A/MS_{error} \sim F_{I-1, IJ(n-1)} \text{ sous } H$$

- Test for effect of factor B

$$H: \beta_j = 0, j = 1, \dots, J$$

- Test statistic :

$$F_B = MS_B/MS_{error} \sim F_{J-1, IJ(n-1)} \text{ sous } H$$

Additive model

- The *additive model* : without interactions
- $y_{ijk} = \mu + \alpha_i + \beta_j + \epsilon_{ijk}$
- $E[\epsilon_{ijk}] = 0$, $Var(\epsilon_{ijk}) = \sigma^2$, $Cov(\epsilon_{ijk}, \epsilon_{i'j'k'}) = 0$ id $(ijk) \neq (i'j'k')$

ANOVA Table

source	df	SS	MS	F
A	$I - 1$	$nJ \sum_{i=1}^I (\bar{y}_{i..} - \bar{y}_{...})^2$	SS_A / df_A	MS_A / MS_{err}
B	$J - 1$	$nI \sum_{j=1}^J (\bar{y}_{.j.} - \bar{y}_{...})^2$	SS_B / df_B	MS_B / MS_{err}
error	$nIJ - I - J + 1$	$\sum_{k=1}^n \sum_{j=1}^J \sum_{i=1}^I (y_{ijk} - \bar{y}_{i..} - \bar{y}_{.j.} + \bar{y}_{...})^2$	SS_{err} / df_{err}	
total (corr.)	$nIJ - 1$	$\sum_{k=1}^n \sum_{j=1}^J \sum_{i=1}^I (y_{ijk} - \bar{y}_{...})^2$		

Indicator variables for the model

- The matrix form for the linear model :

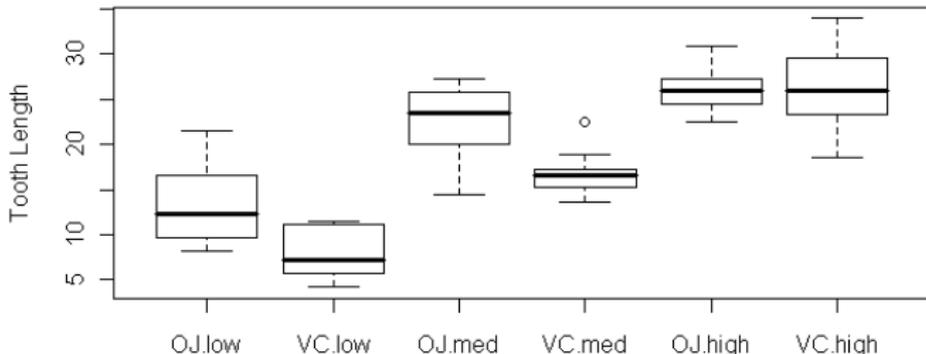
$$\mathbf{Y} = \mathbf{X}\boldsymbol{\beta} + \boldsymbol{\epsilon}$$

- According to the form of the matrix \mathbf{X} , we are in the case of :
 - *linear regression* (\mathbf{X} is then comprised of the constant 1 and p explanatory variables), or
 - *factorial model* (\mathbf{X} is comprised of **indicator variables associated with the levels** of the factor(s))
 - *ancova* (\mathbf{X} is comprised of both qualitative and quantitative variables)

Example : ToothGrowth

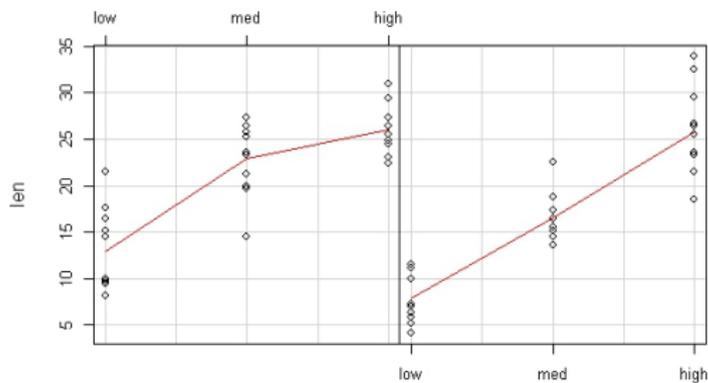
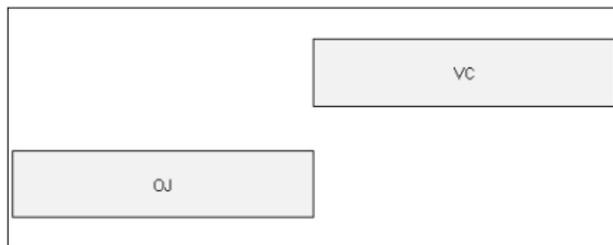
- “The response is the length of odontoblasts (teeth) in each of 10 guinea pigs at each of three dose levels of Vitamin C (0.5, 1, and 2 mg) with each of two delivery methods (orange juice or ascorbic acid).”

Boxplots of Tooth Growth Data



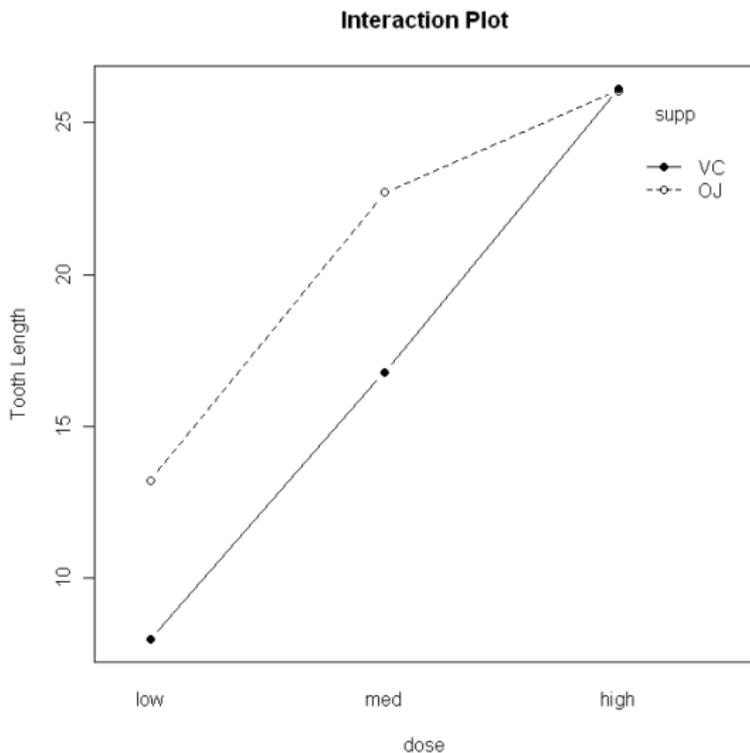
Example, cont : Graphics

Given : supp



ToothGrowth data: length vs dose, given type of supplement

Example, cont. : Interaction plot



Example, cont : ANOVA table output

```
> aov.out = aov(len ~ supp * dose, data=ToothGrowth)
> summary(aov.out)
```

	Df	Sum Sq	Mean Sq	F value	Pr(>F)	
supp	1	205.3	205.3	15.572	0.000231	***
dose	2	2426.4	1213.2	92.000	< 2e-16	***
supp:dose	2	108.3	54.2	4.107	0.021860	*
Residuals	54	712.1	13.2			

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

Unbalanced designs

- When all sample sizes are equal, the main effects and interactions can be estimated independently *independently*
- That is because of the orthogonality of the sub-spaces that correspond to the different model effects
- This is no longer the case when the sample sizes are different (unbalanced case) :
$$SS_{Model} \neq SSA + SSB + SSAB$$
- For an unbalanced design, effect estimation must be *adjusted* (for the other effects in the model) : the estimated values depend on the other terms in the model and their order of entry
- We can no longer carry out tests $F = \frac{MS_x}{MS_{error}}$
- We must carry out sub-model tests

Example, cont : Unbalanced subset

	L	M	H
VC	4.2 11.5 7.3	16.5 16.5 15.2 17.3	23.6 18.5
OJ	15.2 21.5 17.6 9.7	19.7 23.3	25.5 26.4 22.4 24.5

Example, cont. : supp 1st

```
> # full interaction model with
> # supp entering first
>
> fit1 <-
  lm(len ~ supp + doselev + supp:doselev,
      data=toothun)
> anova(fit1)
```

Analysis of Variance Table

Response: len

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
supp	1	174.46	174.46	17.3664	0.0011049
doselev	2	375.75	187.87	18.7012	0.0001495
supp:doselev	2	17.70	8.85	0.8808	0.4377931
Residuals	13	130.60	10.05		

Example, cont : doselev 1st

```
> # full interaction model with doselev
> # entering first
>
> fit2 <-
  lm(len ~ doselev + supp + supp:doselev,
      data=toothun)
> anova(fit2)
```

Analysis of Variance Table

Response: len

	Df	Sum Sq	Mean Sq	F value	Pr(>F)
doselev	2	396.08	198.04	19.7131	0.0001158
supp	1	154.13	154.13	15.3428	0.0017685
doselev:supp	2	17.70	8.85	0.8808	0.4377931
Residuals	13	130.60	10.05		

Summary : numerical and graphical analysis

- Design plot
- Boxplots of outcome for each factor level
- Interaction plots
- Write out model, assumptions, define all parameters
- ANOVA table
- Plots for assumption checking/model assessment
- Example of full analysis at :
<https://www.guru99.com/r-anova-tutorial.html>